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Gene expression profiles of nucleus accumbens, prefrontal cortex and hippocampus in an animal model of schizophrenia: proposed candidate genes

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Introduction: It has been suggested that schizophrenia may be induced by "accidents" or injuries that occur during early brain development and result in a reduction of the neural connections in different regions. In this study, we evaluated differences in the expression of brain genes using a recognized experimental prototype of schizophrenia: the animal model of ventral hippocampal lesion in neonate rats (VHLN) compared to control animals.

Methods: Using microarray technology, we obtained gene expression profiles of three brain areas (nucleus accumbens, prefrontal cortex and hippocampus) of juvenile (45 days) and adult (90 days) Wistar male rats that underwent either VHLN or sham VHLN.

Results: Based on three criteria: 1) expression in more than one brain area, 2) participation in cellular pathways relevant to the central nervous system (CNS), 3) Z-score values >2 (overexpression) and <-2 (underexpression), we found overexpression of the *ppp3cb*, *dctn1*, *jag1*, *ide*, *limk2* and *cpz* genes and underexpression of *chrna4* and *sod1*.

Conclusions: Two of the genes proposed in this paper, *limk2* and *cpz*, have not been previously associated with schizophrenia, so future studies will be necessary to understand their possible role in the pathogenesis of this disease.

Keywords: Schizophrenia, Animal model, Microarrays, Expression, Candidate genes

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Perfiles de expresión génica de núcleo accumbens, corteza prefrontal e hipocampo en un modelo animal de Esquizofrenia: una propuesta de genes candidatos

Introducción: Se ha sugerido que la Esquizofrenia puede ser inducida por "accidentes" o lesiones durante el desarrollo temprano del cerebro del individuo, que conllevan a una reducción en las conexiones neuronales de diferentes regiones. En este trabajo, hemos evaluado las diferencias en la expresión de genes cerebrales, usando un reconocido prototipo experimental para Esquizofrenia: el modelo animal de lesión en hipocampo ventral en ratas neonatas (LHVN), respecto a animales control.

Metodología: Mediante la técnica de chips de ADN, se obtuvieron los perfiles de expresión génica de tres áreas cerebrales (núcleo accumbens, corteza prefrontal e hipocampo) de ratas macho Wistar juveniles (45 días) y adultas (90 días) sometidas o no a LHVN.

Resultados: Con base a tres criterios: 1) expresión en más de un área cerebral, 2) participación en rutas celulares relevantes para el sistema nervioso central (SNC), 3) valores de Z-score >2 (sobre-expresión) y <-2 (sub-expresión); se encontraron sobre-expresados los genes: *ppp3cb*, *dctn1*, *jag1*, *ide*, *limk2* y *cpz*, y sub-expresados: *chrna4* y *sod1*.

Conclusiones: Dos de los genes propuestos en este trabajo: *limk2* y *cpz*, no han sido relacionados previamente con Esquizofrenia, por lo que se hará necesario realizar estudios futuros para dilucidar sus respectivas contribuciones en la etiopatogenia de esta enfermedad.

Palabras clave: Esquizofrenia, Modelo animal, Microarreglos, Expresión, Genes candidatos

INTRODUCTION

Schizophrenia is a serious mental disorder that affects 1 in 100 people worldwide and is characterized by distorted thoughts and perceptions that are probably due to alterations in different neurotransmitter systems^{1,2}. Various hypotheses have been proposed to explain the pathogenesis of this disease, the genetic component being the most important³. However, due to the unavailability of brain tissues from patients and healthy controls for this type of studies, experimental animal models have been used^{4,5}.

The implementation and use of animal models in psychiatry has clear limitations because we are attempting to reproduce complex human behaviors in a healthy animal⁴⁻⁶. Nonetheless, animal models of psychiatric disorders have made it possible to explore the therapeutic potential of specific drugs for treating these disorders, and to obtain relevant data about the mechanisms of action of these drugs; models are also valuable tools for determining the neurobiological substrates of psychiatric disorders⁷. For the study of schizophrenia, several animal models, pharmacological models (using phencyclidines [PCP] and ketamine)⁸⁻¹⁰, genetic models (induced by genetic mutations or deletions)¹¹⁻¹³ and neurodevelopmental models (induced by physical or neurotoxic lesions and environmental factors during neural development) have been proposed¹⁴⁻¹⁶.

It has been suggested that schizophrenia may be due to "injuries" that occur during the early brain development of the individual and reduce neural connections in different limbic regions and the prefrontal cortex¹⁴.

The ventral hippocampal lesion in neonate rats (VHLN) model has been widely tested. In this model, a small excitotoxic lesion is made in the hippocampus of the immature brain of neonate rats and the rats are then allowed to mature further. It has been observed that the injured animals, in the adult stage, exhibit conduct related to the positive and negative symptoms of schizophrenia, such as hyperlocomotion, reduction in prepulse inhibition, memory deficits and decreased social interaction, among other^{15,16}.

DNA microarrays are a valuable genomic tool for studying complex diseases, facilitating the global evaluation of expression of a large number of genes in various tissues and/or physiological conditions¹⁷. Using this methodology, we obtained gene expression profiles at 45 and 90 days for three different brain areas (hippocampus, prefrontal cortex and nucleus accumbens) of male Wistar rats with a neonatal lesion in ventral hippocampus (VHLN). These profiles were compared with those of animals with sham lesions, considered negative controls of the disease, in order to detect significant changes in gene expression that could be related to the schizophrenia phenotype in both juvenile and adult rats.

METHODOLOGY

Biological material

The study began with twenty male Wistar rats from litters of rats inseminated in the laboratory and kept in individual isolation cages under inverted 12-hour light-dark cycle conditions. The total sample was divided into four groups: a) 5 VHLN-juvenile rats, b) 5 sham injured juvenile rats, c) 5 VHLN-adult rats, d) 5 sham injured adult rats. All experiments were performed according to the regulations established by the Mexican official standard for the use and care of laboratory animals "NOM-062-ZOO-1999" and the regulations of the ethics committee of the International Association for the Study of Pain¹⁸.

Newborn rats (10-13 g weight) were injured between days 5 and 7: the pups were randomly assigned to injury with ibotenic acid or sham injury with PBS. The animals were anesthetized using hypothermia by placing them on ice for 10-15 minutes. In order to perform bilateral lesions, the animals were immobilized on a Kopf stereotaxic fixed platform: an incision was made in the scalp and either ibotenic acid (0.15 µl/min for 2 min) (Sigma) or PBS (sham lesion) was administered through a needle according to the following coordinates: anteroposterior (AP) -2.5, mediolateral ±2.5 and dorsoventral (DV) -3.3 in relation to Bregma¹⁹.

The animals were sacrificed by decapitation at 45 days (juvenile group: VHLN and sham) and 90 days (adult group: VHLN and sham). The hippocampus, prefrontal cortex and nucleus accumbens were dissected according to previously standardized procedures²⁰ and frozen at -80°C until RNA extraction.

RNA extraction

RNA was extracted from the 3 previously dissected brain areas using TRIZOL (Life Technologies), according to the manufacturer's instructions.

Microarray design, reading and standardization:

Five thousand *Rattus norvegicus* oligonucleotides from the Oligosets Operon (http://www.operon.com/arrays/oligosets_overview.php) library were used. The microarray design and manufacture was carried out at the Cellular Physiology Unit of the National University of Mexico (UNAM)²¹.

Microarray hybridization started with 10 µg of total RNA, from which dUTP-Cy3 or dUTP-Cy5-labeled cDNA was generated with the CyScribe First-Strand cDNA kit (Amersham). Fluorophore uptake was confirmed by reading the absorbance at 555 nm for Cy3 and at 655 nm for Cy5²¹.

Table 1 The number of genes that did not change expression is shown in the first part. The number of genes that significantly changed expression after normalization is shown in the lower part. (Alma D. Genis-Mendoza)

Juveniles		Adults	
Unchanged genes	%	Unchanged genes	%
4348	86.95	4331	86.61

	JUVENILES					ADULTS				
	Down	%	Up	%	Total (%)	Down	%	Up	%	Total (%)
N. ACCUMBENS	326	6.52	317	6.34	12.86	351	7.02	291	5.82	12.83
PREFRONTAL CORTEX	362	7.24	302	6.04	13.28	372	7.44	310	6.20	13.64
HIPPOCAMPUS	321	6.42	329	6.58	13.00	351	7.02	333	6.66	13.68

Signal and image acquisition and quantification was carried out with ScanArray 4000 equipment and ScanArray 4000 (Packard Biochip) software. All images were captured using 65% PMT gain, 70-75% laser power and 10-micron resolution at a 50% sweep rate. For each point labeled with Cy3 or Cy5, we calculated the average density and mean background value with Array Pro Analyzer (Cybernetics) software²¹. Gene expression was analyzed with the genArise statistical program developed at the Computer Unit of the Cellular Physiology Unit, UNAM (<http://www.ifc.unam.mx/genarise/>) to evaluate the degree of variation in gene expression (z-score). According to this criterion, elements with a z-score value greater or less than two standard deviations are differentially expressed genes²¹.

RESULTS

After standardization, our analysis identified 652 genes that significantly changed their expression, of which 316 were found to be overexpressed (OE) and 336 were underexpressed (UE). The percentage of OE and UE genes for young rats was observed to be almost the same (about 6.5%), whereas the UE gene percentage (7%) was slightly higher than the OE gene percentage (6%) for adult rats. However, the overall percentage of genes that modulated their expression for each tissue remained at $13\% \pm 0.38$ (Table 1). Genes were sorted into 3 groups according to the z-score (>1 and <-1, >2 and <-2, and >3 and <-3). Subsequently, to identify the metabolic and signaling pathways involved in each of the proteins encoded by these genes, we accessed the KEGG metabolic pathways database (<http://www.genome.jp/kegg/kegg2.html>), which is a genomic mapping tool.

Gene group with z-score value >1 and <-1

At this stage, general information was obtained for both overexpressed and underexpressed genes, and a list of all the metabolic pathways for each brain area and the number of genes involved was prepared. We then selected

and plotted the pathways in which a higher number of genes (more than 5 genes) were modulated. Among the most compromised pathways were neuroactive ligand-receptor interaction (>10 genes), calcium signaling (>7 genes) and cancer (>7 genes). It is noteworthy that there were genes whose modulation was significant for two or more pathways.

Group of genes with z-score value >2 and <-2

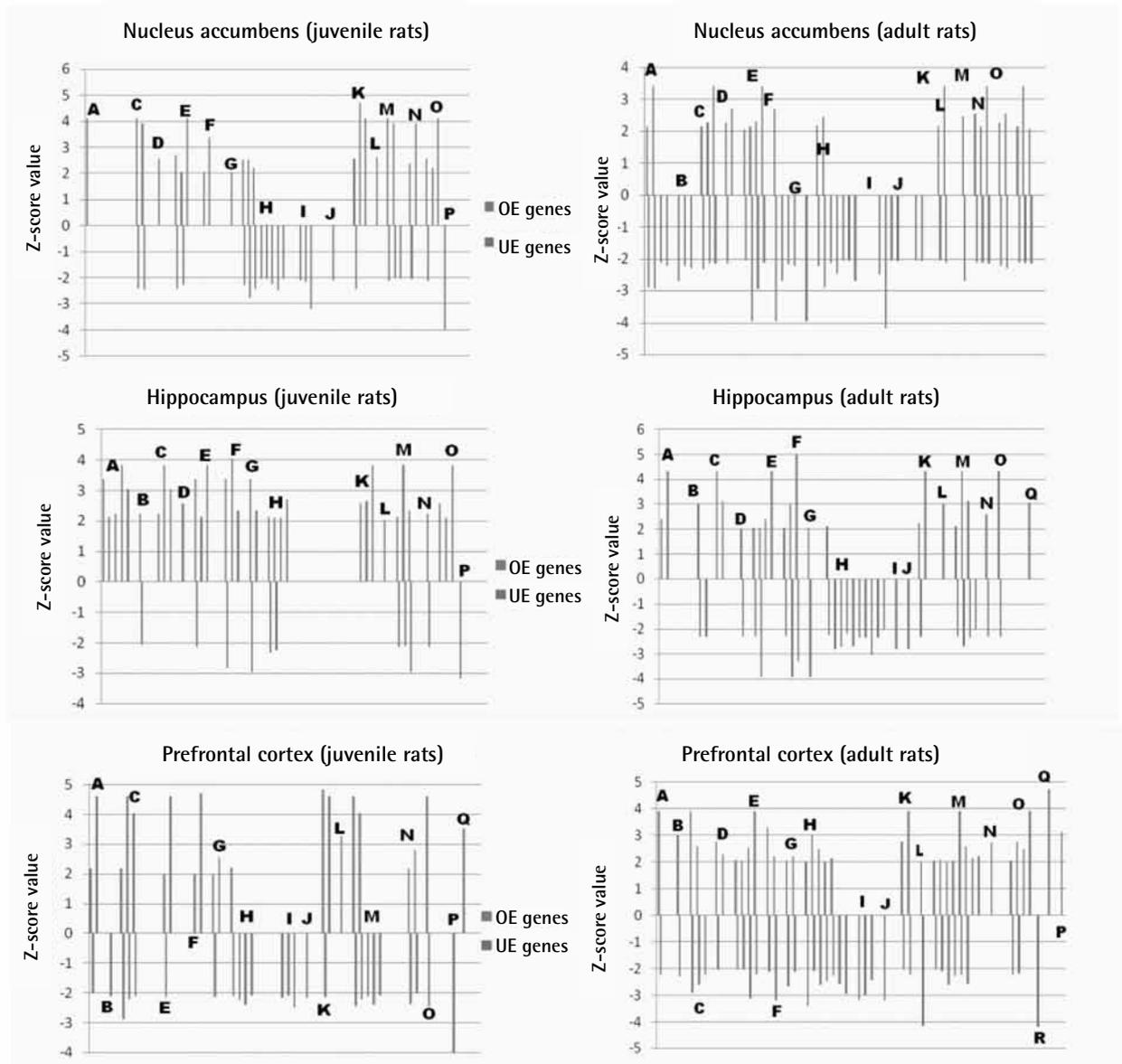
We worked with genes with a z-score value greater than 2 and less than -2 for each tissue for a more robust analysis. Eighteen pathways associated with mental disorders were selected. For each tissue, the metabolic pathways of interest and participating genes were plotted against z-scores (Figure 1).

Because the same gene could be expressed in two or more of the brain areas evaluated (nucleus accumbens, hippocampus and prefrontal cortex), we looked for genes that were expressed in more than one area of interest in order to limit our analysis. As can be seen in Figure 2, in both the juvenile and adult groups, the expression of certain genes is regulated coincidentally in more than one tissue. We found a higher number of underexpressed genes than overexpressed genes, the group of adult rats having the highest number of underexpressed genes. Interestingly, among the overexpressed genes, *ppp3cb* and *dctn1* modulated their expression in all three areas of the brain and in both stages (juvenile and adult).

Group of genes with z-score values >3 and <-3

Lastly, the genes that presented larger changes in expression compared to their control group were analyzed. Similarly, genes that modulated their expression in more than one tissue were detected. Of all of these, *ppp3cb* varied

z-score >2 and <-2

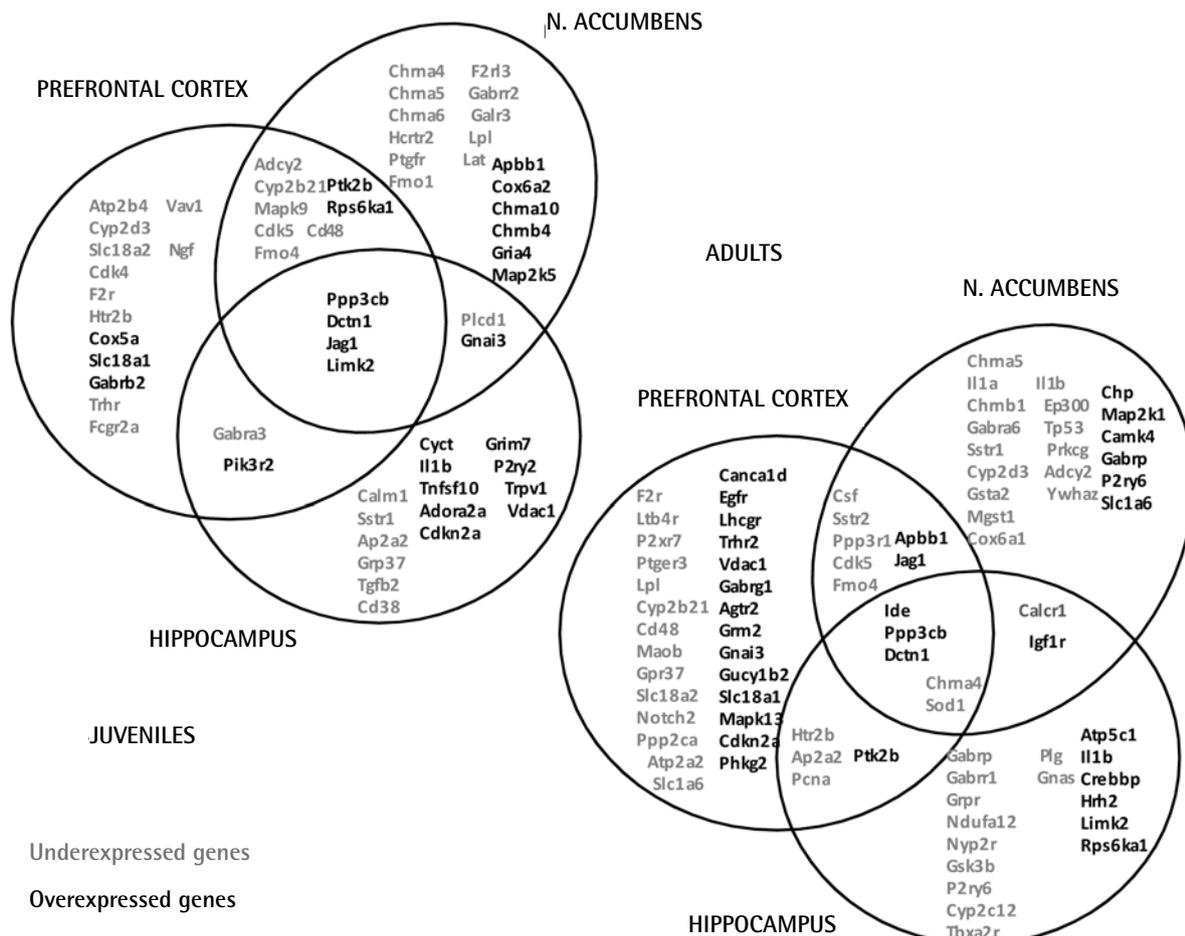


- A) Apoptosis
- B) Cell cycle
- C) NK-mediated cytotoxicity
- D) Long-term depression
- E) Alzheimer
- F) Huntington
- G) Parkinson
- H) Neuroactive ligand-receptor interaction
- I) Cyt-P450 drug metabolism
- J) Cyt-P450 xenobiotic metabolism
- K) Axon orientation

- L) Notch signaling pathway
- M) Calcium signaling pathway
- N) Neurotrophin signaling pathway
- O) Glutamatergic synapsis
- P) Neuronal differentiation and function
- Q) Neuroendocrine response and behavior
- R) Neuroplasticity

Figure 1

The graphs show the number of overexpressed and underexpressed genes involved in pathways of interest in the group of juvenile and adult rats. Each gene is plotted relative to its Z-score value. (Alma D. Genis-Mendoza)



nucleus accumbens (N), prefrontal cortex (P), hippocampus (H)

Figure 2 | This figure shows the candidate genes obtained from the following three criteria: 1) Z-score values >2 and <-2); 2) expression in more than one brain area; and 3) participation in relevant metabolic pathways of the central nervous system

its expression in the three areas of the brain in both groups. In this category, we found another gene with a high z-score, *cpz* 4.37.

Finally, after taking into account z-score values, expression in more than one brain area and participation in metabolic pathways of interest, eight candidate genes were proposed for the schizophrenia study. In Table 2 we summarize the results of the z-score values for the proposed genes; it can be observed that *ppp3cb* and *dctn1* had the highest z-score values in all three areas evaluated.

DISCUSSION

Using microarray technology, we evaluated the expression of the genes in different brain regions using the VHLN animal model for schizophrenia. We worked with 4

experimental groups of Wistar male rats: VHLN juvenile rats, sham-injured juvenile rats, VHLN adult rats and sham-injured adult rats. The analysis was divided into 3 groups according to the z-score value (>1 and <-1, >2 and <-2, and >3 and <-3). Not all the routes observed were directly associated with mental disorders, so we identified 18 pathways relevant to the CNS (genes with z-score >2 and <-2). Finally, we examined the genes with z-score values >3 and <-3 to identify the candidate genes of interest with the greatest variations in expression.

Group of genes with z-scores >1 and <-1

This group included a large number of metabolic and signaling pathways that were found to be differentially altered in the three brain areas. The pathways with the largest number of modulated genes were cancer, neuroactive

Symbol	Name	Description	A Score	P Score	H Score
Limk2	Kinase 2 with LIM domain	Proteins with LIM domains are involved in diverse cell-signaling processes, such as cytoskeletal organization, organogenesis and the cell cycle	4.710	4.834	2.659
Ppp3cb	Phosphatase 3 protein, catalytic subunit, beta isoform	Phosphatase-dependent calmodulin protein; it dephosphorylates nuclear factors of activated T cells and may play a role during skeletal muscle atrophy	4.130	4.600	3.846
Dctn1	Dynactin 1	Dynein microtubule component activated by ATPase, which acts as a microtubule engine	3.378	4.717	4.042
Jag1	Jagged 1	Ligand responsible for activation of the Notch1 receptor. Signaling through <i>Notch</i> is involved in the development of most tissues	2.625	3.254	2.036
Ide	Insulin-degrading enzyme	Enzyme involved in the degradation of bioactive peptides, including insulin, beta-endorphin, atrial natriuretic peptide and beta-amyloid	2.292	2.531	2.052
Sod1	Superoxide dismutase 1, soluble	Catalyzes the conversion from superoxide to hydrogen peroxide and molecular oxygen involved in the oxidative stress response	-2.193	-3.187	-3.301
Chrna4	Cholinergic receptor, nicotinic alpha-4	Belongs to the superfamily of ligand-activated gated ion channels that play a role in the rapid transmission of signals in synapses	-2.909	-3.420	-2.814

ligand-receptor interaction and calcium signaling; the last two pathways were of interest.

Group of genes with z-score >2 and <-2

In this segment, we detected 7 genes of interest whose expression was modified in the three brain areas and had ontologies related to the pathways of interest. The calcium signaling and neuroactive ligand-receptor interaction pathways continued to contribute a large number of differentially expressed genes, whereas neuronal differentiation and function, neuroendocrine response and behavioral pathways had less participation. It should be noted that there were overexpressed and underexpressed genes in most of the pathways, except for the route of drug and xenobiotic metabolism by Cyt-P450, where only underexpressed genes were found (Figure 1).

Group of genes with z-scores >3 and <-3

Finally, this analysis allowed us to propose 8 genes as potential candidates for the genetic approach to schizophrenia. Six of these genes (*ppp3cb*, *dctn1*, *ide*, *limk2*, *jag1* and *cpz*) were overexpressed, while the remaining two (*sod1* and *chrna4*) were underexpressed (Table 2).

Among the genes with the highest z-score values, *ppp3cb* (Table 2) was noteworthy for being overexpressed in the group of juvenile and adult rats. This gene encodes the catalytic subunit of the β isoform of phosphatase 3 protein. This enzyme interacts selectively and noncovalently with calmodulin in response to an increase in intracellular calcium levels²². Recently, a genome study in Taiwanese families with schizophrenia suggested the linkage of locus 10q22.3, where the *anxa7*, *dnajc9*, *zmynd17* and *pppp3cb* genes are located, the same genes associated with schizophrenia and with cognitive and attention deficit problems²³. In addition, a meta-analysis was conducted to evaluate the entire genome of individuals with schizophrenia in various populations. Among the genes proposed as candidate genes was *ppp3cc*, which encodes the gamma isoform of the calcineurin enzyme. Other genes associated were *nos1ap*, *rgs4*, *uhmk1*, *nrg1* and *znf804a*, among others²⁴.

On the other hand, the human *dctn1* gene encodes the major p150 subunit of dynactin, a macromolecular complex consisting of 10 polypeptides, which is required for the retrograde axonal transport of cellular vesicles and organelles through the microtubule system. Numerous studies have shown the role of p150 in psychotic disorders associated with delusions and hallucinations²⁵, and neurodegenerative disorders such as Huntington disease²⁶ and amyotrophic lateral sclerosis (ALS)²⁷.

The differential regulation at the level of *ppp3cb* and *dctn1* transcripts was notable in both the juvenile and adult rats (z-score values >3), which suggests that both may contribute from the onset of the first psychotic symptoms.

Another of the genes that was overexpressed with a high z-score in all three brain areas studied was *ide*, which encodes insulin-degrading enzyme (IDE) (Table 2). This gene is situated in locus 10q23-q25, previously associated with schizophrenia²⁸, and encodes a zinc-dependent metallopeptidase that degrades insulin, glucagon, β -amyloid, β -endorphin, IGF-I and IGF-II. Deficient expression of this enzyme has been associated with Alzheimer disease and type 2 diabetes mellitus^{29,30}. It has also been suggested that haloperidol could have an effect on IDE activity through changes in the level of expression of its substrates²⁸. However, since our data revealed a significant increase in *ide*, it is possible that the regulation of both enzyme levels and activity occurs post-transcriptionally, as described in other members of the metallopeptidase family³¹.

Another transcript expressed abundantly in this study was *jag1*. The resulting protein is the ligand of Notch 1, a transmembrane receptor activated before birth that induces radial glial differentiation³², and that promotes the differentiation of progenitor cells into astroglial scaffolding cells after birth³³. In patients with schizophrenia, it was recently shown that *jag1* expression is significantly increased, exhibiting positive correlation with canonical mediators of the Notch pathway, which influences cell proliferation and the cell cycle, and also participates in oligodendrocyte synthesis^{34,35}. We observed *jag1* overexpression only in the group of juvenile rats. In this respect, we might speculate that *jag1* overexpression could reflect activation of this pathway to promote neurogenesis in response to injury caused by neonatal injury, thus reactivating the neuronal development of the animals.

The *chrna4* gene that encodes a nicotinic acetylcholine receptor belonging to the superfamily of ion channels was found to be underexpressed in the adult animals. As is well-known, this class of receptors plays a major role in the transmission of nerve signals in neuronal synapses. It has been suggested that cholinergic-nicotinic dysfunction may contribute to cognitive impairment in schizophrenia by mediating dopamine release. Furthermore, it has been seen that neurocognitive deficits in schizophrenia can be relieved temporarily by nicotine administration^{36,37}; since then, several nicotinic receptor subtypes have been examined as candidates associated with this disease^{37,38}.

Another potential candidate gene for study in schizophrenia is *sod1*, which we found to be significantly underexpressed in adult rats. The superoxide dismutase 1 protein encoded by this gene is one of two human isoenzymes responsible for the destruction of superoxide free radicals in cells. Mutations in this gene have been associated with familial ALS²⁷. A decrease in superoxide dismutase 1 enzyme

expression may potentiate oxidative stress by increasing the vulnerability of neuronal cells to free radicals, which is consistent with reported brain damage in patients with schizophrenia³⁹.

It was noteworthy that the two underexpressed genes *sod1* and *chrna4* only had decreased levels in the group of adult rats, suggesting that these changes may be associated with positive and negative symptoms of schizophrenia.

Moreover, our findings showed that *limk2* was overexpressed in the juvenile rat group. LIM kinase genes encode a group of protein kinases involved in diverse biological functions. Despite the fact that *limk2* has not been associated with schizophrenia, it has been associated with apoptosis as a protein that protects against ischemia in a rat model of diabetes^{40,29}, and in an animal model of Alzheimer disease, increased *limk2* expression was observed with degeneration of synaptic structures⁴¹. It is known that LIMK2 is phosphorylated and activated by ROCK kinases, thus phosphorylating cofilin, a protein that prevents actin polymerization and favors depolymerization, which are necessary for cell motility⁴². An increase in the expression of *limk2* may be disturbing cytoskeletal dynamics due to failure to regulate actin, which would have negative implications for early neuronal migration in injured juvenile rats and is consistent with our model of schizophrenia.

Finally, in the group of juvenile and adult rats we found overexpression of the carboxypeptidase Z gene (*cpz*). The protein has an N-terminal domain with 30% amino acid identity with the "frizzled" (Fzd) domain present in Wnt-interacting proteins^{43,44}. Carboxypeptidase Z is abundant in the extracellular matrix of the placenta and is found in smaller quantities in brain, lung, thymus and kidney³⁸. This protein had never been associated before with schizophrenia; in contrast, another enzyme of this family, glutamate carboxypeptidase II (GCPII) has been associated with mental disorders, including schizophrenia⁴⁵.

Although the exact function of CPZ is unknown, it is likely that it plays a role in the regulation of embryonic development by interaction with Wnt proteins. Furthermore, it has been reported that CPZ expression is much lower in adult tissues than in embryonic organs⁴⁶. In turn, it has been shown in hippocampal neuron cultures that Wnt-mediated signaling is important for axonal development and guidance⁴⁶, regulating cytoskeletal dynamics in the axonal growth cone and branching⁴⁷, and leads to an increase in the formation and organization of new presynaptic terminals^{48,49}.

Our study showed that the *cpz* gene did not turn off after embryonic development and continued to be overexpressed in injured adult rats. If this protein remains highly expressed in the adult brain, it is very likely that dysregulation of Wnt-mediated signaling exists.

To date, *cpz* and *limk2* genes have never before been associated with schizophrenia and, because of their significant overexpression in our study model, these genes can be expected to play an important role in the development and course of the disorder. Undoubtedly, future research and new experimental strategies should be designed with the aim of clarifying the contribution of LIMK2 and CPZ proteins in this disease and establishing their role as potential biomarkers.

With respect to the three brain areas assessed (nucleus accumbens, hippocampus and prefrontal cortex) in the group of injured animals, a reduction was observed in the number of genes regulated compared with the group of sham injured animals. This is consistent with the cognitive impairment that has been observed in the brains of patients with schizophrenia, where neuronal deterioration and/or damage has been demonstrated in the hippocampus and prefrontal cortex⁵⁰. Given the cognitive impairment manifested by patients with schizophrenia, it is possible that damage to the prefrontal cortex is observed in the long term, which in turn is consistent with the VHLN animal model of schizophrenia, which is based on the neurodevelopmental hypothesis and goes hand-in-hand with an early prenatal brain lesion^{51,52} and the involvement of morphogenesis in the dorsolateral cortex region⁵³. Furthermore, our findings suggest the possibility that the maturation of local inhibitory circuits in the prefrontal cortex may be altered in rats with hippocampal injury, as has been reported in a previous study⁵⁴.

One limitation of this study was that we evaluated gene expression profiles using the DNA 5k chip designed by the Cell Physiology Institute, UNAM. We know that late-generation DNA chips currently available on the market can be used to study more genes and other features; analyses with such chips might identify other genes that we did not observe with this chip. However, we believe that our findings are relevant because the modulation of some genes already associated with schizophrenia was observed, as well as two genes that so far have not been linked to schizophrenia and could play a role in the disease.

CONCLUSIONS

This work showed that the transcriptome of juvenile and adult rats with VHLN was differentially modified compared to the control group. Significant overexpression and underexpression of several genes was reported, some previously associated with schizophrenia and others not. Our contribution in this regard is to propose eight potential candidate genes for analysis in this disorder, two of which have not been previously associated with schizophrenia (*limk2* and *cpz*). These unpublished findings open new questions and broaden the horizons for future research at

the functional level in both animal models and humans, which may allow reliable diagnostic molecular biomarkers to be established in schizophrenia.

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REFERENCES

1. Quintero J, Barbudo del Cura E, López-Ibor MI, López-Ibor JJ. La evolución del concepto de Esquizofrenia Resistente al Tratamiento. *Actas Esp Psiquiatr*. 2011;39(4):236-50.
2. <http://www.imss.gob.mx>
3. Ripke S, Sanders AR, Kendler KS, Levinson DF, Sklar P, Holmans PA, et al. Genome-wide association study identifies five new schizophrenia loci. *Nat Genet*. 2011;43(10):969-76.
4. Ulloa RE, Nicolini H, Fernández-Guasti A. Age differences in an animal model of obsessive-compulsive disorder: participation of dopamine: dopamine in an animal model of OCD. *Pharmacol Biochem Behav*. 2004;78(4):661-6.
5. Ulloa RE, Nicolini H, Fernández-Guasti A. Sex differences on spontaneous alternation in prepubertal rats: implications for an animal model of obsessive-compulsive disorder. *Prog Neuropsychopharmacol Biol Psychiatry*. 2004;28(4):687-92.
6. Fernández-Guasti A, Ulloa RE, Nicolini H. Age differences in the sensitivity to clomipramine in an animal model of obsessive-compulsive disorder. *Psychopharmacology (Berl)*. 2003;166(3):195-201.
7. Ulloa RE, Nicolini H, Avila M, Fernández-Guasti A. Age onset subtypes of obsessive compulsive disorder: differences in clinical response to treatment with clomipramine. *J Child Adolesc Psychopharmacol*. 2007;17(1):85-96.
8. Olszewski RT, Janczura KJ, Ball SR, Madore JC, Lavin KM, Lee JC, et al. NAAG peptidase inhibitors block cognitive deficit induced by MK-801 and motor activation induced by d-amphetamine in animal models of schizophrenia. *Transl Psychiatry*. 2012;3(2):145.
9. Chatterjee M, Verma R, Ganguly S, Palit G. Neurochemical and molecular characterization of ketamine-induced experimental psychosis model in mice. *Neuropharmacology*. 2012;63(6):1161-71.
10. Zuo D, Bzdega T, Olszewski RT, Moffett JR, Neale JH. Effects of N-acetylaspartylglutamate (NAAG) peptidase inhibition on release of glutamate and dopamine in prefrontal cortex and nucleus accumbens in phencyclidine model of schizophrenia. *J Biol Chem*. 2012;287(26):21773-82.
11. Nakajima M, Mori H, Nishikawa C, Tsuruta M, Okuyama S, Furukawa Y. Psychiatric disorder-related abnormal behavior and habenulointerpeduncular pathway defects in Wnt1-cre and Wnt1-GAL4 double transgenic mice. *J Neurochem*. 2013;124(2):241-9.
12. Haque FN, Lipina TV, Roder JC, Wong AH. Social defeat interacts with *Disc1* mutations in the mouse to affect behavior. *Behav Brain Res*. 2012;233(2):337-44.
13. Marongiu MF, Poddie D, Porcu S, Manchinu MF, Castelli MP,

- Sogos V, et al. Reversible disruption of pre-pulse inhibition in hypomorphic-inducible and reversible CB1^{-/-} mice. *PLoS One*. 2012;7(4):e35013.
14. Powell SB. Models of neurodevelopmental abnormalities in schizophrenia. *Curr Top Behav Neurosci*. 2010;4(4):435-81.
 15. Le Pen G, Grottick AJ, Higgins GA, Martin JR, Jenck F, Moreau JL. Spatial and associative learning deficits induced by neonatal excitotoxic hippocampal damage in rats: further evaluation of an animal model of schizophrenia. *Behav Pharmacol*. 2000;11(3-4):257-68.
 16. Albert H.C. Wong, Barbara K. Lipska, Olga Likhodi, Ernie Boffa, Daniel R. Weinberger, James L. Kennedy, et al. Cortical gene expression in the neonatal ventral-hippocampal lesion rat model. *Schizophrenia Research*. 2005;77:261-70
 17. Guido Lastra L, Camila Manrique. Microarreglos: herramienta para el conocimiento de las enfermedades. *Revista colombiana de reumatología*. 2005;12(3):263-7.
 18. Zimmermann M. Ethical principles for the maintenance and use of animals in neuroscience research. *Neurosci Lett*. 1987;73(1):1.
 19. Lipska BK, Jaskiw GE, Chrapusta S, Karoum F, Weinberger DR. Ibotenic acid lesion of the ventral hippocampus differentially affects dopamine and its metabolites in the nucleus accumbens and prefrontal cortex in the rat. *Brain Res*. 1992;585(1-2):1-6.
 20. Mostalac-Preciado CR, de Gortari P, López-Rubalcava C. Antidepressant-like effects of mineralocorticoid but not glucocorticoid antagonists in the lateral septum: interactions with the serotonergic system. *Behav Brain Res*. 2011;30;223(1):88-98.
 21. Pérez-Carreón JI, Martínez-Pérez L, Loredó ML, Yañez-Maldonado L, Velasco-Loyden G, Vidrio-Gómez S, et al. An adenosine derivative compound, IFC305, reverses fibrosis and alters gene expression in a pre-established CCI (4) induced rat cirrhosis. *Int J Biochem Cell Biol*. 2010;42(2):287-96.
 22. Miyakawa T, Leiter LM, Gerber DJ, Gainetdinov RR, Sotnikova TD, Zeng H, et al. Conditional calcineurin exhibit multiple abnormal behaviors related to schizophrenia. *Proc Natl Acad Sci USA*. 2003;100(15):8987-92.
 23. Liu CM, Fann CS, Chen CY, Liu YL, Oyang YJ, Yang WC, et al. ANXA7, PPP3CB, DNAJC9, and ZMYND17 genes at chromosome 10q22 associated with the subgroup of Schizophrenia with deficits in attention and executive function. *Biol Psychiatry*. 2011;70(1):51-8.
 24. Ng MYM, Levinson DF, Faraone SV, Suarez BK, DeLisi LE, Arinami T, et al. Meta-analysis of 32 genome-wide linkage studies of schizophrenia. *Molecular Psychiatry*. 2009;14(8):774-85.
 25. Kurian SM, Le-Niculescu H, Patel SD, Bertram D, Davis J, Dike C. Identification of blood biomarkers for psychosis using convergent functional genomics. *Molecular Psychiatry*. 2011;16:37-58.
 26. Engelender S. Huntingtin-associated protein 1 (HAP1) interacts with the p150Glued subunit of dynactin. *Hum Mol Genet*. 1997;6(13):2205-12.
 27. Shaw PJ. Molecular and cellular pathways of neurodegeneration in motor neuron disease. *Journal of Neurology Neurosurgery and Psychiatry*. 2005;76(8):1046-57.
 28. Bernstein HG, Ernst T, Lendeckel U, Bukowska A, Ansorge S, Stauch R, et al. Reduced neuronal expression of insulin-degrading enzyme in the dorsolateral prefrontal cortex of patients with haloperidol-treated, chronic schizophrenia. *J Psychiatric Res*. 2009;43(13):1095-105.
 29. Wei L, Sun D, Yin Z, Yuan Y, Hwang A, Zhang Y, et al. A PKC-beta inhibitor protects against cardiac microvascular ischemia reperfusion injury in diabetic rats. *Apoptosis*. 2010;15(4):488-98.
 30. Hou Y, Zhou L, Yang QD, Du XP, Li M, Yuan M, et al. Changes in hippocampal synapses and learning-memory abilities in a streptozotocin-treated rat model and intervention by using fasudil hydrochloride. *Neuroscience*. 2012;3:200:120-9.
 31. Toupance S, Brassart B, Rabenoelina F, Ghoneim C, Vallar L, Polette M, et al. Elastin-derived peptides increase invasive capacities of lung cancer cells by post-transcriptional regulation of MMP-2 and uPA. *Clin Exp Metastasis*. 2012;29(5):511-22.
 32. Gaiano N, Nye JS, Fishell G. Radial glial identity is promoted by Notch1 signaling in the murine forebrain. *Neuron*. 2000;26(2):395-404.
 33. Tanigaki K, Nogaki F, Takahashi J, Tashiro K, Kurooka H, Honjo T. Notch1 and Notch3 instructively restrict bFGF-responsive multipotent neural progenitor cells to an astroglial fate. *Neuron*. 2001;29(1):45-55.
 34. Chambers CB, Peng Y, Nguyen H, Gaiano N, Fishell G, Nye JS. Spatiotemporal selectivity of response to Notch1 signals in mammalian forebrain precursors. *Development*. 2001;128(5):689-702.
 35. Kerns D, Vong-Ghe S, Barley K, Dracheva S, Katsel P, Casaccia P, et al. Gene expression abnormalities and oligodendrocyte deficits in the internal capsule in schizophrenia. *Schizophr Res*. 2010;120(1-3):150-8.
 36. López-Valdés H, García-Colunga J. La participación de los receptores de acetilcolina nicotínicos en trastornos del sistema nervioso central. *Salud mental*. 2003;26(3):66-72.
 37. Kishi T, Ikeda M, Kitajima T, Yamanouchi Y, Kinoshita Y, Kawashima K. Genetic association analysis of tagging SNPs in alpha4 and beta2 subunits of neuronal nicotinic acetylcholine receptor genes (CHRNA4 and CHRN2) with schizophrenia in the Japanese population. *Journal of Neural Transmission*. 2008;115(10):1457-6.
 38. De Luca V, Voineskos S, Wong G, Kennedy JL. Genetic interaction between alpha4 and beta2 subunits of high affinity nicotinic receptor: analysis in schizophrenia. *Experimental Brain Research*. 2006;174(2):292-6.
 39. García-Valencia J, Miranda AL, López-Jaramillo CA, Palacio-Acosta CA, Gómez-Franco J, Opsina-Duque J. Esquizofrenia y Neurodesarrollo. *Revista colombiana de psiquiatría*. 2005;34(01):63-76.
 40. te Velthuis Aartjan JW, Bagowski CP. PDZ and LIM domain-encoding genes: molecular interactions and their role in development. *Review Article. The ScientificWorld Journal*. 2007;7:1470-92.
 41. Hou Y, Zhou L, Yang QD, Du XP, Li M, Yuan M, et al. Changes in hippocampal synapses and learning-memory abilities in a streptozotocin-treated rat model and intervention by using fasudilhydrochloride. *Neuroscience*. 2012 3;200:120-9.
 42. Croft D, Crighton D, Samuel M, Lourenco F, Munro J, Wood J, et al. p53-mediated transcriptional regulation and activation of the actin cytoskeleton regulatory RhoC to LIMK2 signaling pathway promotes cell survival. *Cell Res*. 2011;21(4):666-8.
 43. Song L, Fricker LD. Cloning and expression of human carboxypeptidase Z, a novel metalloproteinase. *J Biol Chem*. 1997;272(16):10543-50.
 44. Reznik SE, Fricker LD. Carboxypeptidases from A to Z: implications in embryonic development and Wnt binding. *Cell Mol Life Sci*. 2001;58:1790-1804.
 45. Guilarte TR, Hammoud DA, McGlothan JL, Caffo BS, Foss CA, Kozikowski AP, et al. Dysregulation of glutamate carboxypeptidase II in psychiatric disease. *Schizophr Res*. 2008;99(1-3):324-32.
 46. Zhou CJ, Zhao C, Pleasure SJ. Wnt signaling mutants have decreased dentate granule cell production and radial glial scaffolding abnormalities. *J Neurosci*. 2004;24(1):121-6.

47. Nathan D, Okerlund, Benjamin NR, Cheyette. Synaptic Wnt signaling a contributor to major psychiatric disorders? *J Neurodev Disord.* 2011;3(2):162-74.
48. Varela-Nallar L, Grabowski CP, Alfaro IE, Alvarez AR, Inestrosa NC. Role of the Wnt receptor Frizzled-1 in presynaptic differentiation and function. *Neural Dev.* 2009;2:41.
49. Sahores M, Gibb A, Salinas PC. Frizzled-5, a receptor for the synaptic organizer Wnt7a, regulates activity-mediated synaptogenesis. *Development.* 2010;137(13):2215-25.
50. Harms MP, Wang L, Csernansky JG, Barch DM. Structure-function relationship of working memory activity with hippocampal and prefrontal cortex volumes. *Brain Struct Funct.* 2012;[Epub ahead of print]
51. Hanlon FM, Houck JM, Pyeatt CJ, Lundy SL, Euler MJ, Weisend MP, et al. Bilateral hippocampal dysfunction in schizophrenia. *Neuroimage.* 2011;58(4):1158-68.
52. Lillrank SM, Lipska BK, Weinberger DR. Neurodevelopmental animal models of schizophrenia. *Clin Neurosci.* 1995;3:98-104.
53. Tseng JR, Kang KW, Dandekar M, Yaghoubi S, Lee JH, Christensen JG. Preclinical efficacy of the c-Met inhibitor CE-355621 in a U87 MG mouse xenograft model evaluated by 18F-FDG small-animal PET. *J Nucl Med.* 2008;49:129-34.
54. Tucker DM, Derryberry D, Luu P. Anatomy and Physiology of Human Emotion: Vertical Integration of Brainstem, Limbic, and Cortical Systems. In J. Borod, Ed. *Handbook of the Neuropsychology of Emotion.* New York: Oxford, 2000.